

Patterns of cross-resistance in a multidrug-resistant small-cell lung carcinoma cell line*

S. P. C. Cole

Departments of Oncology and Pharmacology and Toxicology, Queen's University, Kingston, Ontario K7L 3N6, Canada

Received 30 November 1989/Accepted 15 February 1990

Summary. H69AR is a multidrug-resistant human small-cell lung carcinoma cell line that was selected in doxorubicin and has previously been shown to be cross-resistant to a variety of natural-product-type anticancer drugs. H69AR is unlike many other multidrug-resistant cell lines in that it does not overexpress P-glycoprotein. In the present study, the drug sensitivity and cross-resistance patterns of H69AR cells were further characterized. A total of 15 drugs belonging to a number of chemical classes were screened. These compounds included anthracyclines, DNA binders (anthrapyrazoles, benzothiopyranindazoles, and pyrazoloacridines), and lipophilic antifolates. The alkylating agent melphalan and the antimetabolite cytosine arabinofuranoside (Ara-C) were also tested. In general, the drug sensitivity and cross-resistance profiles of H69AR cells were consistent with those reported by others using other drug-resistant cell lines. However, there were several unexpected instances of cross-resistance. Thus, the H69AR cell line was more resistant than its parent cell line to the potent 3'-deamino-3'-(3-cyano-4-morpholinyl) doxorubicin, bisantrene, the pyrazoloacridine PD 114541, Ara-C, and melphalan. In addition, no cross-resistance to the four lipophilic antifolates tested, including trimetrexate, was found. The absence of a consistent pattern among the various drug-resistant cell lines indicates that assumptions about the efficacy of anticancer drugs in multidrug resistance should be made with caution.

flux pump, has been shown to be essential for the expression of the MDR phenotype in many in vitro model systems [4, 12] and appears to be clinically relevant in some malignancies [11, 13, 23, 27, 32]. However, there are some tumour types, such as lung cancers [26], in which P-glycoprotein does not appear to play a clinical role. In addition, there are a number of MDR cell lines that do not overexpress P-glycoprotein, and other mechanisms (e.g., altered topoisomerase II activity, altered intracellular drug distribution, and changes in drug detoxification pathways) have been implicated [7, 20, 28, 29, 37, 42]. One such cell line is the small-cell lung carcinoma cell line, H69AR, derived in this laboratory [29].

Two pharmacological approaches are being followed in efforts to overcome the problem of MDR. One involves the use of agents that enhance the chemosensitivity of known antineoplastic agents [14]. The other approach involves the screening of analogues of drugs in current clinical use as well as novel compounds against resistant cell lines. Numerous in vitro studies of both types have been reported using cell lines that overexpress P-glycoprotein [2, 16, 19, 25, 36, 44, 45] (and others). However, there is limited data with respect to cell lines in which resistance is not mediated by P-glycoprotein [6]. In this paper, the patterns of cross-resistance and drug sensitivity of the non-P-glycoprotein-mediated MDR small-cell lung carcinoma cell line, H69AR [29], are described. This information may be useful in the development of more effective therapeutic strategies in lung cancer and other tumours in which P-glycoprotein is not believed to play a major role in resistance.

Introduction

The development of multidrug resistance (MDR) presents a major obstacle to the successful treatment of many human tumours. P-glycoprotein, a plasma membrane ef-

Materials and methods

Drugs and chemicals. Doxorubicin, 4-demethoxydaunorubicin (idarubicin), and 4'-deoxy-4'-iododoxorubicin were provided by Adria Laboratories (Columbus, Ohio), and 3'-deamino-3'-(3-cyano-4-morpholinyl)doxorubicin (MRA-CN) was provided by Dr. E. M. Acton (NIH, Bethesda, Md.). Bisantrene was a gift of American Cyanamid Co. (Pearl, N. Y.). The anthrapyrazoles [piroxastrone (CI-942), CI-937, and CI-941], the pyrazoloacridines PD 115934 and PD 114541, and the benzothiopyranindazoles PD 114595 and CI-958 (PD 118484) were

* Supported by a grant from the National Cancer Institute of Canada
Correspondence and proofs: S. P. C. Cole, Ph. D., Cancer Research Laboratories, Botterell Hall Rm 331, Queen's University, Kingston, Ontario K7L 3N6 Canada, TEL 613-545-6358, FAX 613-544-9708

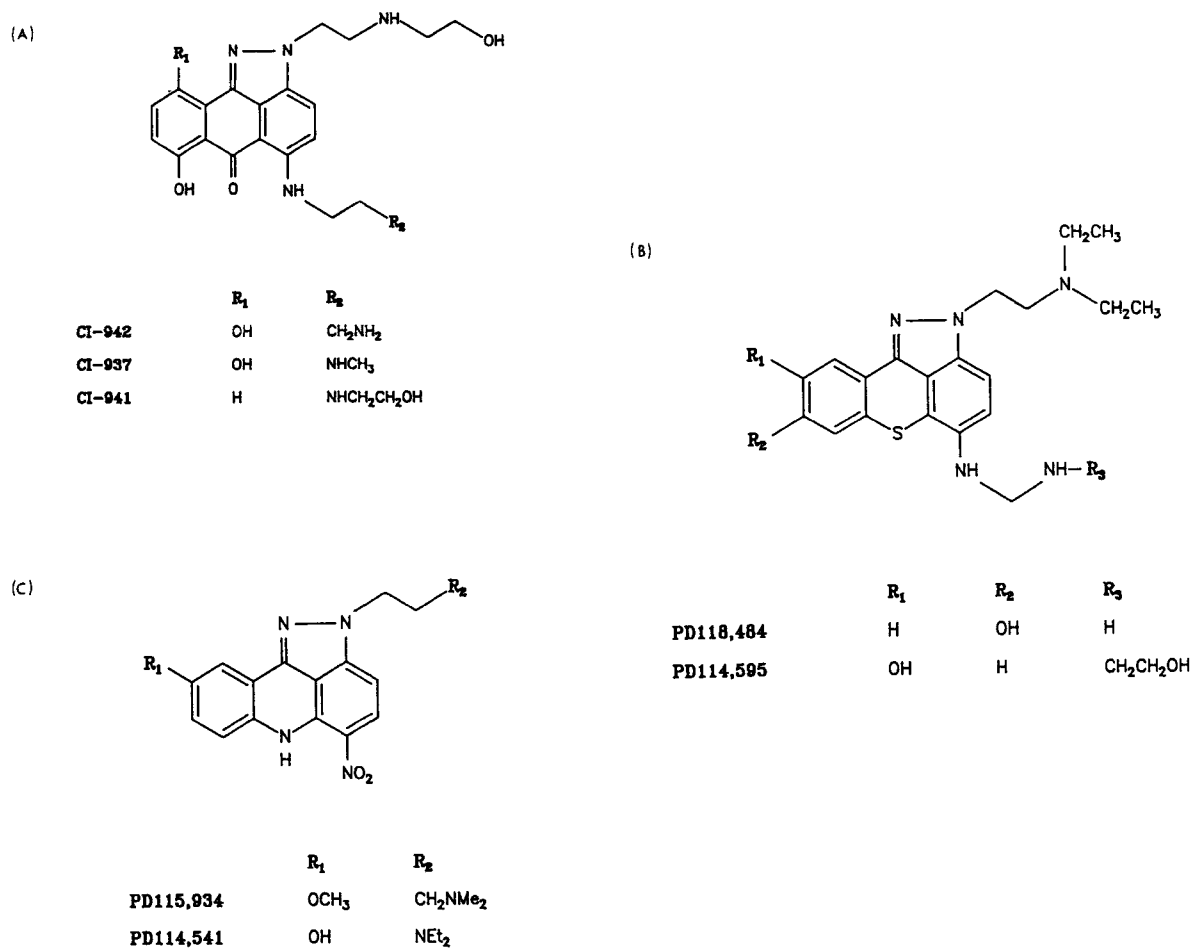


Fig. 1 A–C. Molecular structures of A anthrapyrazoles, B benzothiopyranoindazoles, and C pyrazoloacridines

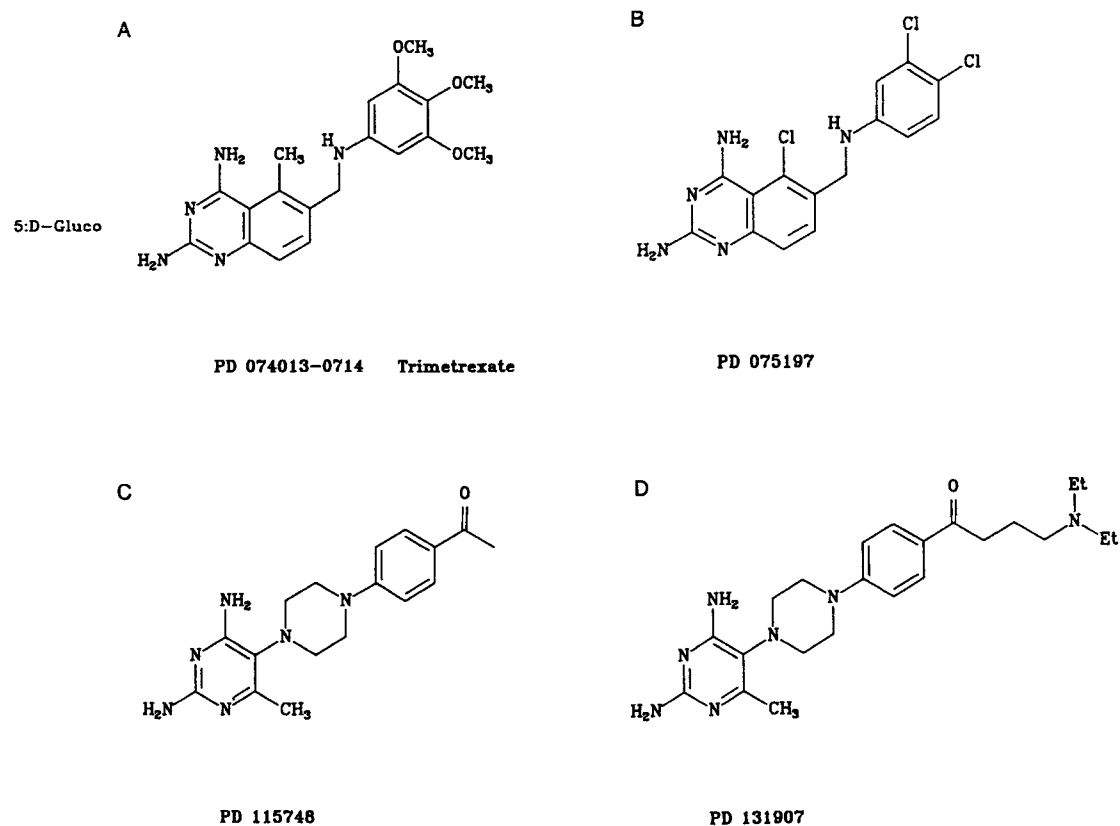


Fig. 2 A–D. Molecular structures of the antifolates A trimetrexate, B PD 75197, C PD 115748, and D PD 131907

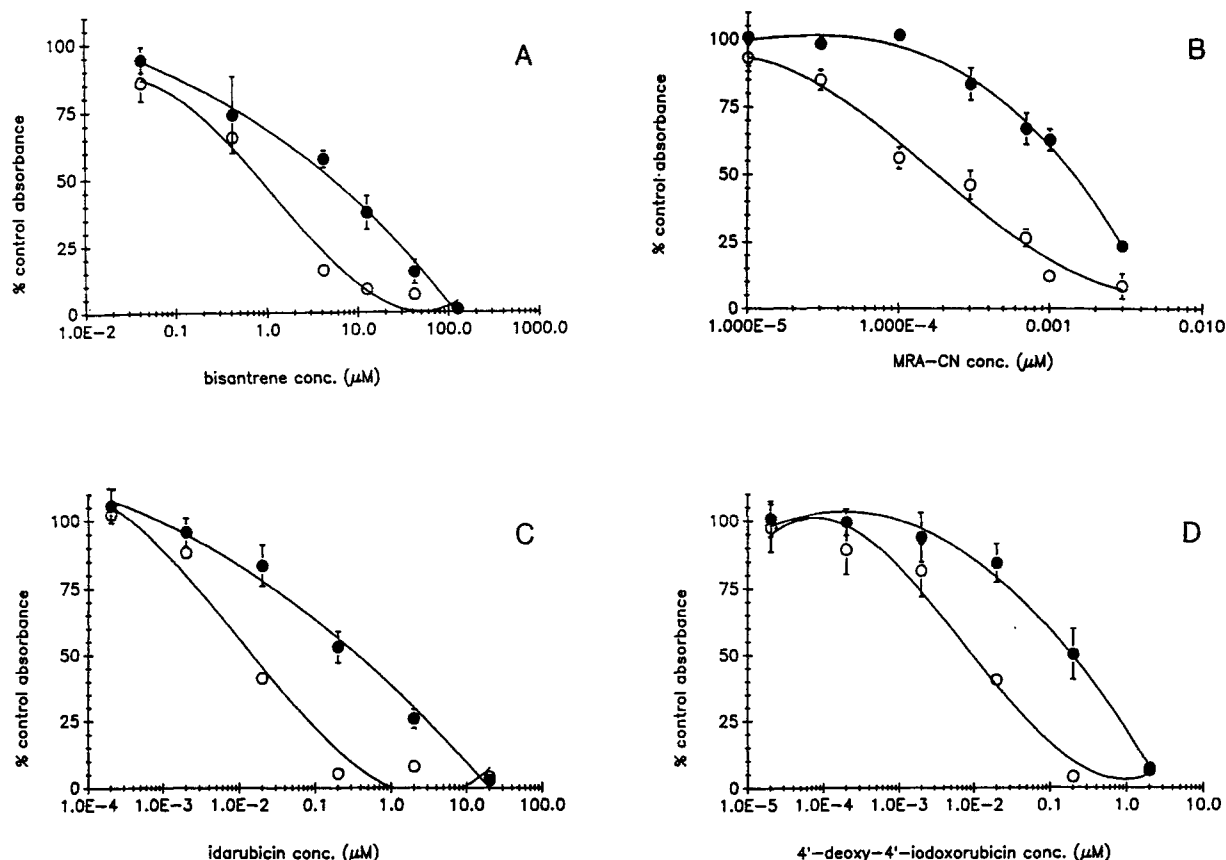


Fig. 3 A–D. Effect of anthracyclines and related compounds on the growth of H69 cells (○) and multidrug-resistant H69AR cells (●), as measured by the MTT assay. The compounds tested included A bisantrene, B MRA-CN, C idarubicin, and D 4'-deoxy-4'-iododoxorubicin. Each point represents the mean \pm SD of four determinations in a typical experiment

Table 1. Cross-resistance of H69AR cells to anthracycline-type compounds

Drug	IC ₅₀ (nM) ^a :		
	H69	H69AR	Resistance ^b (n-fold)
Doxorubicin	74 \pm 45 (n = 28)	4,002 \pm 2,310 ^c (n = 16)	54
Bisantrene	675 \pm 350 (n = 4)	2,901 \pm 1,250 ^c (n = 3)	4
MRA-CN	0.157 \pm 0.061 (n = 3)	0.853 \pm 0.395 ^c (n = 3)	6
Idarubicin	25.6 \pm 10.8 (n = 3)	290 ^c (n = 2)	11
4'-Deoxy-4'-iododoxorubicin	18.0 \pm 6.1 (n = 3)	385 ^c (n = 2)	21

^a Values given represent the mean \pm SD of the results obtained in two or more independent experiments

^b IC₅₀ H69AR: IC₅₀ H69

^c Significantly different from H69 values ($P < 0.05$) according to the unpaired *t*-test

provided by Dr. H. Showalter (Parke-Davis, Ann Arbor, Mich.). The molecular structures of these compounds are shown in Fig. 1. The quinazoline antifolates trimetrexate (PD 74013-714) and PD 75197 and the diaminopyrimidines PD 115748 and PD 131907 were provided by Dr. W. D. Klohs (Parke-Davis). The molecular structures of these compounds are shown in Fig. 2. 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT), cytosine arabinofuranoside (Ara-C), and melphalan were obtained from Sigma Chemical Co. (St. Louis, Mo.).

Cytotoxicity testing. The H69 cell line was provided by J. Minna (NIH, Bethesda, Md.) and its MDR variant, H69AR, has been described previously [29]. These cell lines were cultured in RPMI 1640 medium supplemented with 5% fetal bovine serum (FBS) (Flow Laboratories) and 4 mM L-glutamine at 37°C in a 95% air/5% CO₂ atmosphere and were free of mycoplasma contamination. Cytotoxicity was measured using a modification of the MTT assay, which has previously been described [5, 30]. In this assay, the tetrazolium salt MTT is converted to a blue formazan product by dehydrogenase enzymes that are active only in living cells. H69 and H69AR cells were seeded into microtitre plates at 2.5×10^4 cells/well in a volume of 100 μ l. Drugs were added 24 h later in a 100- μ l volume at twice the desired final concentration. Stock solutions of idarubicin, doxorubicin, MRA-CN, 4'-deoxy-4'-iododoxorubicin, Ara-C, PD 114595, PD 114541, trimetrexate, piroxantrone, CI-937, CI-941, CI-958, and PD 115934 were prepared in tissue-culture medium and diluted as required. PD 75197, PD 115748, and PD 131907 were dissolved in dimethyl sulfoxide and diluted in RPMI 1640/5% FBS as required. Melphalan was dissolved in acidified isopropanol and bisantrene, in acidified medium prior to dilution in RPMI 1640/5% FBS. After a 4-day exposure to drug, 100 μ l medium was removed, 25 μ l MTT (2 mg/ml PBS) was added, and the plates were incubated for 3 h at 37°C. The resulting formazan crystals were solubilized by the addition of 100 μ l isopropanol: 1 N HCl (24:1), thorough mixing, and further in-

Table 2. Cross-resistance of H69AR cells to anthrapyrazoles, benzothioipyranoidazoles and pyrazoloacridines

Compound ^a	Experiment	IC ₅₀ (μM) ^b :		Resistance ^c (<i>n</i> -fold)
		H69	H69AR	
Anthrapyrazoles:				
CI-942	1	5.9	>10	>1.7
(pirovantrone)	2	1.9	>10	>5.3
CI-937	1	0.20	2.1	11
	2	0.22	2.4	11
CI-941	1	0.050	0.55	11
	2	0.065	0.81	13
Benzothioipyranoidazoles:				
PD 114595		0.042 ± 0.013 (<i>n</i> = 3)	1.068 ± 0.623 ^d (<i>n</i> = 4)	25
CI-958 (PD 118484)		0.44 ± 0.19 (<i>n</i> = 3)	1.22 ± 0.30 ^d (<i>n</i> = 3)	2.8
Pyrazoloacridines:				
PD 115934		0.35 ± 0.07 (<i>n</i> = 3)	0.91 ± 0.09 ^e (<i>n</i> = 3)	2.6
PD 114541		0.028 ± 0.006 (<i>n</i> = 3)	6.13 ± 3.30 ^d (<i>n</i> = 3)	219

^a Molecular structures of these compounds are shown in Fig. 1

^b Where fewer than three experiments were done or it was not possible to calculate an IC₅₀ because the percentage of control absorbance did not fall below 50% over the range of concentrations tested, the results of individual experiments are given. Otherwise, the results shown represent the mean ± SD of results obtained in three or more independent experiments

^c IC₅₀ H69AR : IC₅₀H69

^d Significantly different from H69 values (*P* < 0.05) according to the unpaired *t*-test

^e Significantly different from H69 values (*P* < 0.01) according to the unpaired *t*-test

incubation for 1 h at 37°C. The plates were scanned at 570 nm with a Dynatech MR600 microtitre plate reader. Within each experiment, determinations were done in quadruplicate and each drug was tested in two or more separate experiments. In addition, a doxorubicin dose-response curve was included as an internal control for each day's experiments. Controls consisted of wells with no cells and wells with cells plus vehicle (baseline absorbance). Results were expressed as a percentage of the baseline absorbance at 570 nm and the IC₅₀ was defined as being the dose of drug that reduced this absorbance to 50% of control values. Statistical analysis was done using an unpaired *t*-test, with a significance level of 0.05.

Results

The cytotoxic effects of four anthracycline-type compounds were determined in the first series of experiments. Typical dose-response curves for bisantrene, MRA-CN, idarubicin, and 4'-deoxy-4'-iododoxorubicin are shown in Fig. 3; the IC₅₀s for these compounds are summarized in Table 1. H69AR cells exhibited a significant cross-resistance to all four compounds. MRA-CN had the highest activity, and cytotoxicity to both the H69 and H69AR cell lines was observed at concentrations of <1 nM. The activities of the two anthracyclines idarubicin and 4'-deoxy-4'-iododoxorubicin were similar and moderately greater than that of doxorubicin. Bisantrene was less toxic than doxorubicin to H69 cells but the two drugs showed similar toxicity to H69AR cells.

In the second series of experiments, the cytotoxic effects of seven DNA-binding compounds were tested and the results are summarized in Table 2. Where IC₅₀ values were obtained, the relative resistance of H69AR cells to these agents ranged from 2- to 219-fold. The three an-

thrapyrazoles varied considerably in their toxicity to H69 cells: pirovantrone (CI-942) was the least active, CI-937 showed intermediate cytotoxicity, and CI-941 was the most toxic. The two benzothioipyranoidazoles displayed about a 10-fold difference in their activity against H69 cells but showed similar toxicity to H69AR cells. This resulted in a modest cross-resistance of H69AR cells to CI-958 (2.8-fold) as compared with PD 114595 (25-fold). The two pyrazoloacridines also varied about 10-fold in their activity against H69 cells, whereas the relative cytotoxicity of these two compounds to H69AR cells was reversed. These differences resulted in a large relative resistance of 219-fold for PD 114541 and a modest relative resistance of 2.6-fold for PD 115934.

In the third group of experiments, four antifolates (Fig. 2) were evaluated for their cytotoxic effects; the results of these experiments are presented in Table 3. Only one of the four compounds, the diaminopyridine PD 115748, exhibited significant differential cytotoxicity to H69 and H69AR cells. In this case, however, the H69 cells were more resistant to PD 115748 than were H69AR cells, giving a ratio of IC₅₀ H69AR : IC₅₀ H69 amounting to 0.3. It should be noted that there was considerable difficulty in getting this compound completely into solution at a concentration of dimethylsulfoxide that did not itself cause toxicity.

In the last series of experiments, melphalan and Ara-C were tested; typical dose-response curves are shown in Fig. 4. The IC₅₀ of Ara-C on H69 cells was 0.589 ± 0.442 μM (*n* = 4). The dose-response curve for Ara-C on H69AR cells was very flat (Fig. 4A), and in most experiments the IC₅₀ was not achieved by the maximal concentration of drug tested (0.5 mM). Thus, it was not

Table 3. Cross-resistance of H69AR cells to trimetrexate and other antifolate compounds

Drug ^b	IC ₅₀ (μM) ^a :		Ratio ^c
	H69	H69AR	
Trimetrexate (PD 074013-0714)	0.025 ± 0.012	0.039 ± 0.027 ^d	1.6
PD 075197	0.068 ± 0.062	0.065 ± 0.114 ^d	1.0
PD 115748	1.244 ± 0.425	0.334 ± 0.394 ^e	0.3
PD 131907	0.376 ± 0.376	0.139 ± 0.176 ^d	0.4

^a Values given represent the mean ± SD of results obtained in three or more independent experiments

^b Molecular structures are given in Fig. 2

^c IC₅₀ H69AR : IC₅₀ H69

^d Not significantly different from H69 values ($P > 0.1$)

^e Significantly different from H69 values ($P < 0.05$)

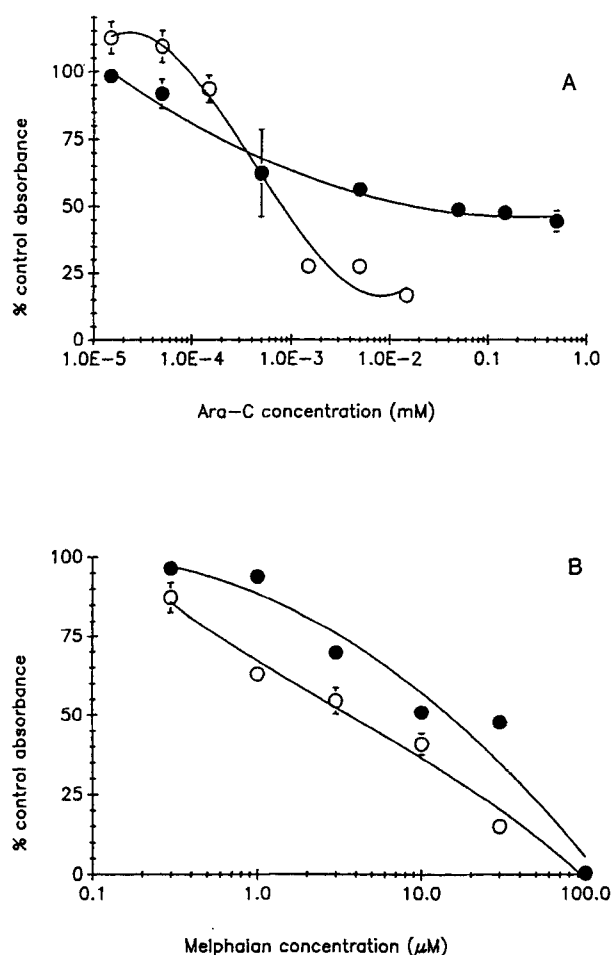


Fig. 4 A, B. Effect of A Ara-C and B melphalan on the growth of H69 cells (○) and multidrug-resistant H69AR cells (●), as measured by the MTT assay. Each point represents the mean ± SD of quadruplicate determinations in a typical experiment

possible to determine accurately the extent of cross-resistance to Ara-C. H69AR cells displayed a 2-fold resistance to melphalan (IC₅₀ H69AR, 14.54 ± 5.99 μM, vs IC₅₀ H69, 7.28 ± 3.08 μM ($n = 5$) that was statistically significant ($P < 0.05$) (Fig. 4B).

Discussion

The present study was undertaken to evaluate the activity of a panel of antineoplastic agents belonging to a number of different chemical classes against the H69AR cell line. This MDR cell line was selected in doxorubicin and does not overexpress P-glycoprotein. Nevertheless, it is cross-resistant to a variety of natural products or their semi-synthetic derivatives, including daunomycin, epirubicin, menogaril, and mitoxantrone, as well as to acivicin, etoposide, gramicidin D, colchicine, and the *Vinca* alkaloids vincristine and vinblastine [29].

The use of doxorubicin in the practice of clinical oncology is often restricted by a cumulative dose-limiting cardiotoxicity as well as by the development of resistance. These limitations have provided the stimulus for research efforts that have resulted in the synthesis of a vast number of anthracycline analogues. Cell lines that overexpress P-glycoprotein have been found to display cross-resistance to most of these analogues, although in many cases this resistance is somewhat diminished as compared with that to doxorubicin [8, 19, 21, 33, 45]. Nevertheless, there are some analogues, most notably, 9-alkyl [8, 19, 34, 45] and morpholinyl derivatives such as MRA-CN [1, 43], which have retained their activity against resistant cell lines to a large degree. As in previous reports with other cell lines [37, 40], MRA-CN was found to be considerably more active than doxorubicin against the H69 and H69AR cell lines (Table 1). However, in contrast to the findings of other investigators, who observed no (or minimal) cross-resistance to MRA-CN [37, 40, 43], the H69AR cells still displayed a 6-fold resistance to this compound (Fig. 3B, Table 1). 4'-Deoxy-4'-iododoxorubicin and idarubicin have shown reduced cross-resistance in other model systems [3, 8, 9, 19, 33], and similar results were found with H69 and H69AR cells (Table 1). Thus, the three anthracyclines tested in the present study, including the highly active MRA-CN, all showed differential toxicity to the sensitive and resistant small-cell lung carcinoma cell lines, albeit to a lesser degree than doxorubicin (Fig. 3C, D; Table 1). A mitoxantrone-selected, drug-resistant colon carcinoma cell line that does not overexpress P-glycoprotein has been found to be cross-resistant to doxorubicin but not to vinblastine or bisantrene [10]. By contrast, H69AR cells are cross-resistant to both the *Vinca* alkaloids [29] and bisantrene (Table 1, Fig. 3A).

The anthrapyrazoles (Fig. 1A) are modified anthracenediones that were developed in an effort to identify DNA-binding antitumour agents with a lower potential for cardiotoxicity than the anthracyclines [18, 38, 39]. In agreement with our findings (Table 2), Klohs et al. [25] observed that piroxantrone was considerably less cytotoxic than doxorubicin to murine P388 cells. These investigators also found that the P388R cell line, which overexpresses P-glycoprotein, displayed cross-resistance to piroxantrone and CI-937, as does the H69AR cell line (Table 2). H69AR cells were cross-resistant to the anthrapyrazole CI-941, as has been reported for other MDR cell lines [8]. The related benzothiopyranindazoles (Fig. 1B) have been tested in two rodent MDR cell lines (Dr. W. Klohs, personal communication). Cross-resistance to both CI-958 (PD 118484)

and PD 114595 was observed in one cell line, whereas in the other, cross-resistance was seen only with CI-958. In all cases, the level of resistance was moderate (≤ 5 -fold). With respect to H69AR cells, the cross-resistance to CI-958 was similar in the rodent cell lines but resistance to PD 114595 was much higher (25-fold); the structural basis for this difference in activity of the two benzothioapyranoin-dazoles is unknown. The pyrazoloacridines (Fig. 1C) have been reported to have selective activity against cell lines derived from solid tumours in vitro [35]. In a study using three MDR cell lines, Sebolt et al. [36] found no cross-resistance to PD 115934 in vitro; similarly, the cross-resistance of H69AR cells to PD 115934 was slight (2.6-fold) (Table 2). In contrast, H69AR cells displayed considerable cross-resistance to PD 114541 (219-fold), as compared with the low level of cross-resistance (2- to 3-fold) observed in the Sebolt et al. study [36]; the structural basis for this difference in activity against H69AR cells shown by the two pyrazoloacridines is also unknown.

Analogues to the antifolate methotrexate have been developed in an attempt to overcome the resistance that frequently occurs after exposure to this drug. One approach has been to synthesize analogues of greater lipophilicity than the parent compound, which can be expected to get into the cell by diffusion rather than by the methotrexate transport system [41]. Although these agents have been effective in circumventing methotrexate resistance [22], cross-resistance to trimetrexate (Fig. 2A) and some other lipophilic antifolates has been observed in several MDR cell lines that overexpress P-glycoprotein [2, 24, 31]. In contrast, significant cross-resistance to trimetrexate was not observed in H69AR cells (Table 3). The other quinazoline antifolate tested, PD 75197 (Fig. 2B), was also equally toxic to H69 and H69AR cells. It is noteworthy that cross-resistance to PD 75197 was not observed in the murine MDR cell line that did exhibit cross-resistance to trimetrexate (Dr. W. Klohs, personal communication). The two diaminopyrimidine antifolates PD 115748 and PD 131907 (Fig. 2C, D), were generally less cytotoxic than the quinazolines, and no cross-resistance in H69AR cells was observed; in contrast, a collateral sensitivity to these two agents was noted, although this effect was significant only in the case of PD 115748 (Table 3). These findings contrast with those reported in a P388 MDR cell line in which significant cross-resistance (111-fold) was observed with PD 131907 but not PD 115748 (Dr. W. Klohs, personal communication). The molecular basis for the differing activities of these four antifolates in the different MDR model systems is unclear at the present time.

Cell lines that possess the MDR phenotype rarely display cross-resistance to antimetabolites and alkylating agents [12, 17, 37, 42]. It is therefore of interest that H69AR cells are cross-resistant to Ara-C and melphalan (Fig. 4). Resistance to the latter drug has been associated with an increase in intracellular glutathione [15]. Since we have found that glutathione levels are diminished in H69AR cells [7], the modest (2-fold) but significant cross-resistance to this agent was particularly surprising.

In summary, we show that many features of the cross-resistance patterns of the H69AR cell line are similar to those obtained in other studies that have used MDR cell

lines that overexpress P-glycoprotein. On the other hand, there were a number of unanticipated observations, viz., the cross-resistance to MRA-CN, bisantrene, PD 114541, ara-C, and melphalan. In addition, the complete lack of cross-resistance to the four lipophilic antifolates was unexpected. Taken together with results from other studies, our findings clearly illustrate the difficulty of drawing general conclusions about the efficacy of anticancer drugs, even within a given chemical class, against MDR cell lines. The basis of the variable cross-resistance patterns of different MDR cell lines is unknown and is probably complex. It is possible that the patterns of cross-resistance are related to the mechanism(s) underlying the resistance phenotype and, therefore, H69AR cells should not be expected to behave like cell lines that overexpress P-glycoprotein. Careful studies with cell lines in which it is known that only a *single* mechanism of resistance is functional are necessary to provide the data that would support or refute this idea. It is also possible that the intrinsic properties of tumours derived from a particular tissue might influence the cross-resistance profiles. In this regard, it would be of interest to determine cross-resistance patterns of a panel of MDR cell lines derived from the same tumour type.

Acknowledgements. The author wishes to thank Dr. Wayne Klohs for helpful discussions. The expert technical assistance of Debra Clements is gratefully acknowledged. Prof. R. Whitney is to be thanked for the preparation of some of the figures.

References

1. Acton EM, Tong GL, Mosher CW, Wolgemuth RL (1984) Intensely potent morpholinyl anthracyclines. *J Med Chem* 27: 638
2. Assaraf YG, Molina A, Schimke RT (1989) Cross-resistance to the lipid-soluble antifolate trimetrexate in human carcinoma cells with the multidrug-resistant phenotype. *J Natl Cancer Inst* 81: 290
3. Barbieri B, Giuliani FC, Bordini T, Casazza AM, Geroni C, Bellini O, Suarato A, Gioia B, Penco S, Arcamone F (1987) Chemical and biological characterization of 4'-iodo-4'-deoxydoxorubicin. *Cancer Res* 47: 4001
4. Bradley G, Juranka PF, Ling V (1988) Mechanism of multidrug resistance. *Biochim Biophys Acta* 948: 87
5. Cole SPC (1986) Rapid chemosensitivity testing of human lung tumor cells using the MTT assay. *Cancer Chemother Pharmacol* 17: 259
6. Cole SPC, Downes HF, Slovak ML (1989) Effect of calcium antagonists on the chemosensitivity of two multidrug resistant human tumour cell lines which do not overexpress P-glycoprotein. *Br J Cancer* 59: 42
7. Cole SPC, Downes HF, Mirski SEL, Clements DJ (1990) Alterations in glutathione and glutathione-related enzymes in a multidrug resistant small cell lung cancer cell line. *Mol Pharmacol* 37: 192
8. Coley HM, Twentyman PR, Workman P (1989) Identification of anthracyclines and related agents that retain preferential activity over Adriamycin in multidrug-resistant cell lines, and further resistance modification by verapamil and cyclosporin A. *Cancer Chemother Pharmacol* 24: 284
9. Dalton WS, Durie BGM, Alberts DS, Gerlach JH, Cress AE (1986) Characterization of a new drug-resistant human myeloma cell line that expresses P-glycoprotein. *Cancer Res* 46: 5125
10. Dalton WS, Cress AE, Alberts DS, Trent JM (1988) Cytogenetic and phenotypic analysis of a human colon carcinoma cell line resistant to mitoxantrone. *Cancer Res* 48: 1882

11. Dalton WS, Grogan TM, Rybski JA, Scheper RJ, Richter L, Kailey J, Broxterman HJ, Pinedo HM, Salmon SE (1989) Immunohistochemical detection and quantitation of P-glycoprotein in multiple drug-resistant human myeloma cells: association with level of drug resistance and drug accumulation. *Blood* 73: 747
12. Gerlach JH, Karter N, Bell DR, Ling V (1986) Multidrug resistance. *Cancer Surv* 5: 25
13. Goldstein LJ, Galski H, Fojo A, Willingham M, Lai S-L, Gazdar A, Pirker R, Green A, Crist W, Brodeur GM, Lieber M, Cossman J, Gottesman MM, Pastan I (1989) Expression of a multidrug resistance gene in human cancers. *J Natl Cancer Inst* 81: 116
14. Gottesman MM, Pastan I (1989) Clinical trials of agents that reverse multidrug resistance. *J Clin Oncol* 7: 409
15. Green JA, Vistica DT, Young RC, Hamilton TC, Rogan AM, Ozols RF (1984) Potentiation of melphalan cytotoxicity in human ovarian cancer cell lines by glutathione depletion. *Cancer Res* 44: 5427
16. Gupta RS, Murray W, Gupta R (1988) Cross resistance pattern towards anticancer drugs of a human carcinoma multidrug-resistant cell line. *Br J Cancer* 58: 441
17. Harker WG, Sikic BI (1985) Multidrug (pleiotropic) resistance in doxorubicin-selected variants of the human sarcoma cell line MES-SA. *Cancer Res* 45: 4091
18. Hartley JA, Reszka K, Zuo ET, Wilson WD, Morgan AR, Lown JW (1988) Characteristics of the interaction of anthracycline anticancer agents with deoxyribonucleic acids: structural requirements for DNA binding, intercalation, and photosensitization. *Mol Pharmacol* 33: 265
19. Hill BT, Dennis LY, Li X-T, Whelan RDH (1985) Identification of anthracycline analogues with enhanced cytotoxicity and lack of cross-resistance to Adriamycin using a series of mammalian cell lines in vitro. *Cancer Chemother Pharmacol* 14: 194
20. Hindenburg AA, Gervasoni JE Jr, Krishna S, Stewart VJ, Rosado M, Lutzky J, Bhalla K, Baker MA, Taub RN (1989) Intracellular distribution and pharmacokinetics of daunorubicin in anthracycline-sensitive and -resistant HL-60 cells. *Cancer Res* 49: 4607
21. Johnson RK, Ovejera AA, Goldin A (1976) Activity of anthracyclines against an Adriamycin (NSC-123127)-resistant subline of P388 leukemia with special emphasis on cinerubin A (NSC-18334). *Cancer Treat Rep* 60: 99
22. Kamen BA, Eibl B, Cashmore A, Bertino J (1984) Uptake and efficacy of trimetrexate (TMQ, 2,4-diamino-5-methyl-6-[3,4,5-trimethoxyanilino)methyl]quinazoline), a non-classical antifolate in methotrexate-resistant leukemia cells in vitro. *Biochem Pharmacol* 33: 1697
23. Kanamaru H, Kakehi Y, Yoshida O, Nakanishi S, Pastan I, Gottesman MM (1989) MDR1 RNA levels in human renal cell carcinomas: correlation with grade and prediction of reversal of doxorubicin resistance by quinidine in tumor explants. *J Natl Cancer Inst* 81: 844
24. Klohs WD, Steinkampf RW, Besserer JA, Fry DW (1986) Cross resistance of pleiotropically drug resistant P388 leukemia cells to the lipophilic antifolates trimetrexate and BW 301U. *Cancer Lett* 31: 253
25. Klohs WD, Steinkampf RW, Havlick MJ, Jackson RC (1986) Resistance to anthracyclines and anthracyclines in multidrug-resistant P388 murine leukemia cells: reversal by calcium blockers and calmodulin antagonists. *Cancer Res* 46: 4352
26. Lai S-L, Goldstein LJ, Gottesman MM, Pastan I, Tsai C-M, Johnson BE, Mulshine JL, Ihde DC, Kayser K, Gazdar AF (1989) MDR1 gene expression in lung cancer. *J Natl Cancer Inst* 81: 1144
27. Ma DD, Davey RA, Harman DH, Isbister JP, Scurr RD, Mackertich SM, Dowden G, Bell DR (1987) Detection of a multidrug resistant phenotype in acute non-lymphoblastic leukemia. *Lancet* i: 135
28. McGrath T, Center MS (1987) Adriamycin resistance in HL60 cells in the absence of detectable P-glycoprotein. *Biochem Biophys Res Commun* 145: 1171
29. Mirski SEL, Gerlach JH, Cole SPC (1987) Multidrug resistance in a human small cell lung cancer line selected in Adriamycin. *Cancer Res* 47: 2594
30. Mosmann T (1983) Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods* 65: 55
31. Ramu A, Fridkin M, Steinherz R (1985) Cross-resistance to esters of methotrexate in a doxorubicin-resistant subline of P388 murine leukemia. *Cancer Chemother Pharmacol* 15: 31
32. Salmon SE, Grogan TM, Miller T, Scheper R, Dalton WS (1989) Prediction of doxorubicin resistance in vitro in myeloma, lymphoma, and breast cancer by P-glycoprotein staining. *J Natl Cancer Inst* 81: 696
33. Schott B, Robert J (1989) Comparative cytotoxicity, DNA synthesis inhibition and drug incorporation of eight anthracyclines in a model of doxorubicin-sensitive and -resistant rat glioblastoma cells. *Biochem Pharmacol* 38: 167
34. Scott CA, Westmacott D, Broadhurst MJ, Thomas GJ, Hall MJ (1986) 9-Alkyl anthracyclines. Absence of cross-resistance to Adriamycin in human and murine cell cultures. *Br J Cancer* 53: 595
35. Sebolt JS, Scavone SV, Pinter CD, Hamelhele KL, Von Hoff DD, Jackson RC (1987) Pyrazoloacridines, a new class of anticancer agents with selectivity against solid tumors in vitro. *Cancer Res* 47: 4299
36. Sebolt J, Havlick M, Hamelhele K, Nelson J, Leopold W, Jackson R (1989) Activity of pyrazoloacridines against multidrug-resistant tumor cells. *Cancer Chemother Pharmacol* 24: 219
37. Seneviratne C, Goldenberg GJ (1989) Further characterization of drug-sensitivity and cross-resistance profiles of cloned cell lines of Adriamycin-sensitive and -resistant P388 leukemia. *Cancer Commun* 1: 21
38. Showalter HDH, Johnson JL, Werbel LM, Leopold WR, Jackson RC, Elslager EF (1984) 5-[(Aminoalkyl)amino]-substituted anthra[1,9-*cd*]pyrazol-6(2H)-ones as novel anticancer agents. Synthesis and biological evaluation. *J Med Chem* 27: 253
39. Showalter HDH, Johnson JL, Hoftiezer JM, Turner WR, Werbel LM, Leopold WR, Shillis JL, Jackson RC, Elslager EF (1987) Anthracycline anticancer agents. Synthesis and structure-activity relationships against murine leukemias. *J Med Chem* 30: 121
40. Sikic BI, Ehsan MN, Harker WG, Friend NF, Brown BW, Newman RA, Hacker MP, Acton EM (1985) Dissociation of antitumor potency from anthracycline cardiotoxicity in a doxorubicin analog. *Science* 228: 1544
41. Sirotak FM, Moccio DM, Kelleher LE, Goutas LJ (1981) Relative frequency and kinetic properties of transport-defective phenotypes among methotrexate-resistant L1210 clonal cell lines derived in vivo. *Cancer Res* 41: 4447
42. Slovak ML, Hoeltge GA, Dalton WS, Trent JM (1988) Pharmacological and biological evidence for differing mechanisms of doxorubicin resistance in two human tumor cell lines. *Cancer Res* 48: 2793
43. Streeter DG, Johl JS, Gordon GR, Peters JH (1986) Uptake and retention of morpholinyl anthracyclines by Adriamycin-sensitive and -resistant P388 cells. *Cancer Chemother Pharmacol* 16: 247
44. Twentyman PR (1988) Modification of cytotoxic drug resistance by non-immuno-suppressive cyclosporins. *Br J Cancer* 57: 254
45. Twentyman PR, Fox NE, Wright KA, Workman P, Broadhurst MJ, Martin JA, Bleehen NM (1986) The in vitro effects and cross-resistance patterns of some novel anthracyclines. *Br J Cancer* 53: 585